



TOXICOKINETICS OF AFLATOXIN B1

AFB₁ absorption, distribution and elimination is rapid in all animal species. Animal studies have shown that under normal conditions, 50-80% of the orally administered dose of AFB₁ is quickly absorbed from the duodenal region of the small intestine of non-ruminant animals (AFSSA, 2009). In ruminants, it passes rapidly through the rumen wall. After absorption toxin enters the liver through the hepatic portal blood supply, where it is metabolised in several derivatives. It is also metabolised in other tissues such as kidney, the intestinal tract and lung, but to a lesser degree (Leong et al., 2012). The flora and fauna in the gastrointestinal tract can also metabolise aflatoxins (Upadhaya et al., 2009).

The susceptibility of an animal to AFB₁ toxicity and carcinogenicity is to a large extent dependent on the rate of metabolism and the type of metabolites that are produced. In most animals, AFB₁ is metabolised to AFP₁, AFM₁, AFQ₁ and AFB₁-8,9-epoxide (Guengerich, 2003; Kamdem et al., 2006). The major route of excretion of AFB₁ and its metabolites is the biliary pathway, followed by the urinary pathway (Gratz, 2007).



The parent compound can also be excreted in faeces. In cows, pigs and sheep, AFM1 is the main unconjugated metabolite in urine. In mammals, the exposure-based biomarker AFM1 can be detected in milk within 12 h after the ingestion of AFB1 (Battacone et al., 2012). The carry-over of AFB1 in feed metabolised into AFM1 in milk for dairy cows is usually 1-2% for low-yielding cows (<30 kg milk yield/day) and up to 6% for high-yielding cows (Veldman et al., 1992).



AFB1, AFM1 and AFP1 can form nucleic acid adducts or undergo conjugation to glutathione (GSH), conversion to dihydrodiols, or binding to serum proteins or other macromolecules (mechanism-based biomarkers). The binding of AFB1 metabolites to proteins and nucleic acids occurs soon after absorption. Both the DNA and the protein adducts have proven useful as both exposure and mechanism-based biomarkers in humans and animals (Guarisco et al., 2008). Around 80% of the depurinated adducts of AFB1 to nucleic acid are excreted within 48 hours of dosing. The correlation between dietary intake and adducts in urine and serum is more variable (IARC, 2002) compared to the close correlation between the levels of adducts in urine and levels in liver (IARC, 1993). Some metabolites bound to nucleic acids can persist in tissues for relatively long periods of time. The urinary DNA adduct reflects recent exposure, as excretion occurs over 24-48 hours, whereas the serum albumin adduct reflects accumulated longer term exposure (2-3 months).

Aflatoxins cause characteristic histopathology of the liver and these lesions are a [effect-based] biomarker of aflatoxicosis (Coppock et al., 2012). The lesions induced by aflatoxins vary with the level and duration of exposure (Coppock et al., 1989). The histopathology observed includes bile duct proliferation (hyperplasia), hepatocyte necrosis, and early fibrosis of the liver (Newberne and Butler, 1969). Species differences occur in the specific histopathology of aflatoxins. Hyperplasia of the bile duct cells occurs rapidly in ducklings and may be present in horses, dogs and chickens, and mild bile duct hyperplasia may be seen in cattle and pigs (Coppock et al., 1989).



References

1. Agence Française de Sécurité Sanitaire des Aliments. Évaluation des risques liés à la présence de mycotoxines dans les chaînes alimentaires humaine et animale; Agence Française de Sécurité Sanitaire des Aliments: Maisons-Alfort, France, 2009; pp. 1-308
2. Baldwin T.T., Riley R.T., Zitomer N.C., Voss K.A., Coulombe R.A. Jr, Pestka J.J., Williams D.E., Glenn A.E. (2011), The current state of mycotoxin biomarker development in humans and animals and the potential for application to plant systems. *World Mycotoxin J* 4:257-270
3. Battacone, G.; Nudda, A.; Rassu, S.P.G.; Decandia, M.; Pulina, G., Excretion pattern of aflatoxin M1 in milk of goats fed a single dose of aflatoxin B1. *J. Dairy Sci.* 2012, 95, 2656-2661
4. Coppock, R.W., Christian, R.G., and Jacobsen. B.J. 2012, Aflatoxins. In *Veterinary Toxicology: Basic and Clinical Principles*. R.C. Gupta, ed. Academic Press/Elsevier, Amsterdam, in press
5. Coppock R.W., Reynolds R.D. and Buck W.B. (1989), Acute aflatoxicosis in feeder pigs resulting from improper storage of corn. *J. Am. Vet. Med. Assoc.*, 195: 1380-1381
6. Gratz (2007), Aflatoxin Binding by Probiotics, Experimental Studies on Intestinal Aflatoxin Transport, Metabolism and Toxicity, Doctoral Thesis; University of Kuopio, Finland
7. Guengerich F.P. (2003), Cytochrome P450 oxidations in the generation of reactive electrophiles: epoxidation and related reactions, *Arch Biochem Biophys*, 409, 59-71
8. Guarisco J.A., Hall J.O., and Coulombe R.A. Jr (2008), Butylated hydroxytoluene chemoprevention of aflatoxicosis - effects on aflatoxin B(1) bioavailability, hepatic DNA adduct formation, and biliary excretion, *Food Chem Toxicol*, 46, 3727-31
9. IARC (1993), IARC Monographs on the Evaluation of Carcinogenic Risks of Chemicals to Humans, Vol. 56, Toxins derived from *Fusarium moniliforme*: Fumonisin B1 and B2 and fusarin C. International Agency for Research on Cancer, Lyon, France
10. IARC (2002), IARC Monographs on the Evaluation of Carcinogenic Risks of Chemicals to Humans, Vol. 82, Some Traditional Herbal Medicines, Some

Mycotoxins, Naphthalene and Styrene, International Agency for Research on Cancer, Lyon, France

11. Kamdem L.K., Meineke I., Godtel-Armbrust U., Brockmoller J., and Wojnowski L. (2006), Dominant contribution of P450 3A4 to the hepatic carcinogenic activation of aflatoxin B1, *Chem Res Toxicol*, 19, 577-86
12. Leong Y.H.(1), Latiff A.A., Ahmad N.I., Rosma A. 2012, Exposure measurement of aflatoxins and aflatoxin metabolites in human body fluids. A short review. *Mycotoxin Res.* 2012 May;28(2):79-87. doi: 10.1007/s12550-012-0129-8. Epub 2012 Apr 3
13. Upadhaya, S.D., Sung, H.G., Lee, C.H., Lee, S.Y., Kim, S.W., Cho, K.J., Ha, J.K. 2009, Comparative study on the aflatoxin B1 degradation ability of rumen fluid from Holstein steers and Korean native goats. *J Vet Sci.* 2009 Mar; 10(1): 29-34
14. Veldman, A.; Meijs, J.A.C.; Borggreve, J.; Heeres-van der Tol, J.J., Carry-over of aflatoxin from cows' food to milk. *Anim. Prod.* 1992, 55, 163-168